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Identification and structural elucidation of ozonation transformation products of estrone

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Abstract

Background: Quantitative methods for the analysis of contaminants of emerging concern (CECs) are abundant in the scientific literature. However, there are few reports on systematic methods of identification and structural identification of transformation products. For this reason, a new method based on high-resolution mass spectrometry and differential analysis was developed in order to facilitate and accelerate the process of identification and structural elucidation of transformation products CECs. This method was applied to the study of ozonation transformation products (OTPs) of the natural hormone estrone (E1).

Results: A control compare trend experiment consisting in the comparison of a control sample to several samples having been exposed to decreasing concentrations of $O_{3(aq)}$ indicated that 593 peaks could be associated with OTPs. After applying various filters to remove background noise, sample contaminants and signal spikes, this data set was reduced to 16 candidate peaks. By inspection of the shape of these peaks, only two compounds OTP-276 (m/z 275.12930) and OTP-318 (m/z 317.14008) were considered as good candidates for further study. Multi-stage tandem mass spectrometry (MS^n) experiments of SPE extracts of the ozonated samples of E1 and of a deuterium-labeled analogue (E1- d_4) showed that OTP-276 and OTP-318 had carboxylic acid and hydroxyl functional groups, as previously reported for OTPs of other hormones. Structures for these two compounds were proposed based on their MS^n spectra.

Conclusion: These results indicate that the method proposed is a systematic and rapid approach to study transformation products of CECs.

Keywords: Transformation products, Differential analysis, Estrone, High-resolution mass spectrometry, Pharmaceuticals, Ozonation

Background

In the past 15 years, the presence and fate of contaminants of emerging concern (CECs) in the environment has been studied extensively. However, relatively less attention has been given to transformation products (TPs) [1]. TPs of CECs can be found in wastewater treatment plants or in the environment as a result of a multitude of abiotic and biotic factors (such as hydrolysis [2], photolysis [3], oxidation [4] and microbial metabolism [5]) acting on parent compounds. TPs are of environmental concern particularly if they are biologically active or resistant to biodegradation. Conservation of biological activity after transformation is plausible if the pharmacophore of the parent compound is preserved [6] and resistance to biodegradation can result after the formation of new functional groups capable

of inhibiting microbial metabolism [7]. Recently, an international expert workshop concluded that the assessment of risks of metabolites and environmental transformation products of pharmaceutical and personal care products is among the top 20 key issues that need to be addressed by the research community [8].

For these reasons, researchers have started to investigate the formation of TPs and have proposed molecular structures based on mass spectrometry [9], especially TPs formed after ozonation [10] which is one of the most studied tertiary water treatments. Different approaches have been used in order to identify and elucidate the structure of ozonation transformation products (OTPs). Huber, et al. [11] used model compounds having the same reactive groups towards O_3 as 17α -ethinylestradiol in order to simplify the identification process of the OTPs of this estrogen. Structural elucidation was performed with gas chromatography–mass spectrometry (GC-MS) and liquid

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chromatography-tandem mass spectrometry (LC-MS/MS) both at low resolution. Bila, et al. [12] compared the retention time and GC-MS spectra of the OTPs of 17 β -estradiol with those of commercial pure standards that were predicted as potential OTPs, based on the molecular structure of the hormone and known O₃ chemistry. The authors also used spectral libraries to identify potential matches. Radjenović, et al. [13], de Oliveira Pereira, et al. [14] and Larcher, et al. [15] employed LC coupled to a hybrid high-resolution mass spectrometer to separate, identify and assign structures to OTPs. Benner and Ternes [16] based the structural elucidation of the OTPs of propranolol on product ion fragmentation trees generated by LC coupled to multi-stage tandem mass spectrometry (MSⁿ). In spite of these studies, there are still few publications in relation to the development of systematic methods of identification and structural elucidation of TPs. Kern, et al. [17] used a database to screen 1794 possible TPs of 52 organic contaminants in preconcentrated extracts of surface waters analyzed by high-resolution mass spectrometry (HRMS). The list was generated by computer prediction of potential microbial TPs as well as experimentally known TPs of 52 organic contaminants. Applying this method the authors were able to identify 19 TPs in the samples. Helbling, et al. [18] used LC-HRMS and data-dependent MS/MS experiments with target and non-target post acquisition processing and found five new TPs formed after microbial degradation of pharmaceuticals. Gómez-Ramos, et al. [19] also employed an accurate mass database of organic contaminants and their more abundant product ions. Their approach was successful to identify eight TPs of organic contaminants in effluent wastewater samples.

The primary objective of this study was to develop a systematic and faster method for the identification and structural elucidation of OTPs of CECs, using HRMS and differential analysis. This method was designed to automatize the identification process and reduce the time required for data interpretation and manual analysis of the large data files generated by HRMS. Identification of OTPs was based on a control compare trend experiment. This experiment consists in the comparison to a control of several samples in which a variable has been modified, in this case the concentration of O₃. A similar approach using differential global profiling was reported previously, however it only compares two sets of samples: a control and a treated group [20]. The advantage of the control compare trend experiment for the identification of OTPs is the possibility of monitoring the signal of the studied CEC and its OTPs as a function of a variable known to affect the formation of OTPs. This approach facilitates the identification of the optimal concentration for the generation of OTPs and, by applying various data filters, the list of OTPs candidates can be narrowed down to a manageable number of peaks.

Once the main OTPs of the target compound are identified, their structure can be elucidated by infusing preconcentrated ozonated samples directly into the mass spectrometer in order to perform MSⁿ experiments at high resolution.

To demonstrate the applicability of this approach, the hormone estrone (E1) was chosen as a model compound and its TPs after ozonation were investigated. As a naturally occurring steroid hormone, E1 is commonly excreted from the body in its active form and has been reported in municipal wastewater effluents at concentrations up to 96 ng L⁻¹ in Canada [21]. Since wastewater treatment plants were not designed to remove this hormone and other CECs, E1 is continuously released via wastewater effluent into surface water bodies and levels as high as 112 ng L⁻¹ have been reported in surface water in the USA [22]. These receiving streams may become sources of raw drinking water or indirectly contribute to contamination of other sources of drinking water sources, as suggested by the frequency of detection of E1 (in 79% of samples) in sources of drinking water in the United States [23]. E1 has been observed to be almost completely removed during wastewater disinfection using ozone [24]. However, bench-scale studies investigating the fate and transformation of E1 during ozonation have revealed the formation of OTPs [11,14].

Materials and methods

Ozonation of estrone samples

Estrone (E1) of high purity (99.0%) was purchased from Sigma-Aldrich. Ozone [O_{3(g)}] was produced in-situ from O_{2(g)} using an Ozomax OZO4VTT generator (Granby, QC) and introduced in pure H₂O using a gas-washing bottle in order to produce an aqueous ozone solution. The concentration of O_{3(aq)} in the solution was determined by means of the Beer-Lambert law by measuring the absorbance at 260 nm and a molar absorptivity value of 3300 M⁻¹ cm⁻¹ [25]. Aliquots of this O_{3(aq)} stock solution (60–982 μ L) were added to 9 mL of a standard solution of E1 in pure H₂O (0.59 mg L⁻¹, free pH with an initial value of 6.05 \pm 0.05) using gas tight syringes according to specific E1:O₃ molar ratios (1:0, 2:1, 1:1, 1:2, 1:5, 1:8). Therefore, applied ozone doses were in the range 0.05 mg L⁻¹ to 0.84 mg L⁻¹. According to Paraskeva, et al. [26], O₃ doses of 4–10 mg L⁻¹ are usually used in WWTPs for disinfection. However preliminary experiments showed that with the conditions used here, O₃ doses of 1 mg L⁻¹ were sufficient to remove E1 below the quantification limit (65 μ g L⁻¹). After addition of O_{3(aq)} the sample volume was brought to 10 mL by addition of pure water. Then samples were covered and stirred for 10 min.

Liquid chromatography-high resolution mass spectrometry analysis of estrone

Initial and residual E1 concentration were measured by liquid chromatography-high resolution mass spectrometry

(LC-HRMS) using an Accela LC system coupled to a hybrid mass spectrometer consisting in a linear ion trap interfaced to an orbital trap (LTQ Orbitrap XL), all manufactured by Thermo Fisher Scientific (Waltham, MA). LC separation was done using a stationary phase consisting of a Hypersil Gold column (50 × 2.1 mm, 1.9 μm) protected with a in-line Direct-Connection UHPLC 0.2 μm filter. All solvents used for mobile phase preparation were Optima LCMS grade and were manufactured by Fisher Scientific. The mobile phase was constituted of H₂O (solvent A) and MeCN (solvent B). LC gradient was the following (%B): 0 min (10%), 1 min (10%), 6 min (80%), 6.1 min (97%), 7.6 min (97%). Re-equilibration time was 2.4 min. Ionization in the negative mode was chosen over ionization in the positive mode since previous studies [14,27] on the OTPs of estrogens have shown that the latter gave better results. Also preliminary experiments (results not shown) indicated better signals of OTPs of E1 with atmospheric pressure chemical ionization (APCI) than with electrospray ionization. For these reasons, the Ion Max API source was used with the APCI probe in the negative mode. APCI(-) source parameters were: vaporizer temperature = 400°C; sheath gas flow = 32 arbitrary units, auxiliary sheath gas flow = 5 arbitrary units; discharge current = 4 μA; capillary temperature = 250°C, capillary voltage = 30 V, tube lens = 70 V. Detection was performed in the orbital trap portion of the mass spectrometer at high resolution ($R_{FWHM} = 41\ 000$ at m/z 269). Scanning was performed from m/z 50 to 350. The full scan cutoff at m/z 350 was set based on the estimation that a change in the molecular mass $> + 80$ u was unlikely for an OTP of E1. For these settings mass accuracy was < 1.8 mmu for m/z 269.

Identification of the main ozonation transformation products of estrone

Since a multitude of substances (solvents, air and glassware contaminants) can be ionized and detected in the full scan mode [28], it is necessary to use a software that can discriminate background signals from the OTPs of E1 present in the samples. To facilitate and accelerate the elucidation process a differential analysis software, Sieve 1.3 developed by Thermo Scientific, was employed. The control compare trend experiment in Sieve was selected in order to monitor the various peaks, which might be associated with TPs, as a function of O₃ dosage. In this type of experiment a sample is designed as control to which others samples are compared and normalized. The sample E1:O₃ 1:8 was selected here as control in the trend experiment knowing that any potential OTP of E1 would have, in this sample, an average intensity > 0 (otherwise the calculated ratio sample to control would return an error). Software settings were the following: frames = up to 1000 with signal threshold > 500

000; m/z start = 200; m/z stop = 350; m/z width = 0.02; Retention time start = 0 min; Retention time stop = 7.5 min; Retention time width = 1 min. Maximum number of frames and intensity threshold were set to limit the identification process to only major OTPs. Also the lower threshold for the m/z range of the frames was set in order to identify the OTPs most closely related to E1 and that may conserve some of its biological activity.

Preconcentration and purification

In order to obtain higher amounts of the suspect OTPs of E1, the experiments described in *Ozonation of estrone samples* section were repeated using 250 mL rather than 10 mL. A semi-batch reactor was employed to produce the O_{3(aq)} solution [29]. The concentration of E1 was also increased to 1.3 mg L⁻¹, corresponding to its solubility limit in H₂O, 1.30 ± 0.08 mg L⁻¹ [30]. In order to obtain a maximum amount of the OTPs the E1:O₃ ratio was set to 1:10. Ozonated samples were preconcentrated using solid-phase extraction (SPE) based on a method described previously [31]. Briefly, since it has been reported that many of the OTPs of E1 have carboxylic acid functions [11,14], ozonated samples were acidified to pH 2.1 with HCl 1 M in order to increase their SPE recovery with Waters Oasis HLB cartridges (6 mL, 150 mg). The cartridges were conditioned with 2 × 2.5 mL MeOH and 2 × 3 mL acidified H₂O with HCl to pH 2.0. Samples were loaded into the cartridges in a Supelco 12-position manifold (St. Louis, MO) at flow rate of 1–4 mL min⁻¹. After loading, cartridges were rinsed with 5 mL of H₂O to wash out any salts and vacuum-dried at 0.68 bar for 5 min. Cartridges were eluted with 2 × 1.25 mL of MeOH and 2 × 1.25 mL of MeCN. Extracts were then evaporated to dryness using a Savant SPD 131DDA Speed Vac Concentrator connected to a RVT4104 refrigerated vapor trap, both manufactured by Thermo Fisher Scientific (Waltham, MA). Finally samples were reconstituted in 50% H₂O-50% MeCN (v/v) and passed through Millipore Millex-LG syringe filters (filter diameter 4 mm, pore size 0.22 μm) before direct infusion into the mass spectrometer to perform MSⁿ experiments.

Structural elucidation and confirmation of the main ozonation transformation products of estrone

Once the main OTPs of E1 were identified, aliquots of the preconcentrated ozonated samples containing the OTPs of interest were introduced in the LC-HRMS system by infusion using a glass syringe. The precursor ions of the OTPs of E1 were then isolated and multiple MSⁿ ($n=2-4$) experiments were performed on the product ions using collision-induced dissociation (CID) and higher energy collision-induced dissociation (HCD). The mass-to-charge ratio (m/z) of the product ions was measured at high resolving power measured at full width at

half-maximum ($R_{FWHM} \approx 190\ 000$ at m/z 231). For these settings mass accuracy was < 0.9 mmu for m/z 231. Elemental formulas were generated from the exact masses of only the frames that were identified as potential OTPs using the differential analysis software and using the "Elemental composition" function in QualBrowser program of Xcalibur 2.1. The elements in use were: C (0 to 18 atoms), O (0–10 atoms) and H (0–40 atoms). Mass tolerance was set to 10 mmu. Ring and double-bond equivalents (RDBE) value was set to -0.5 to 20 while charge was set to -1 . Structures were proposed based on known fragmentation pathways for negative even-electron ions [32] as well as basic principles of the stability of the product ions and the neutral products [33].

In order to confirm the structure of the identified OTPs of E1, a deuterated E1 standard, estrone-2,4,16,16- d_4 (99%) was bought from CDN isotopes (Point-Claire,

QC). The E1- d_4 standard used had two deuterium atoms (^2H , but for simplicity D will be used instead) in the aromatic ring, in C-2 and C-4, and two geminal D atoms in C-16 (Figure 1). Semi-batch ozonation experiments, as described in section Preconcentration and purification, were performed with this standard to generate the same OTPs (but with different mass) in order to confirm the sites on the E1 molecule where reaction with O_3 took place and propose structures of the MS^n product ions of the identified OTPs of E1.

Results and discussion

Identification of estrone ozonation transformation products by differential analysis

Preliminary experiments with the acquisition files showed that identification of OTPs of E1 based on present knowledge of O_3 chemistry and previously reported OTPs was

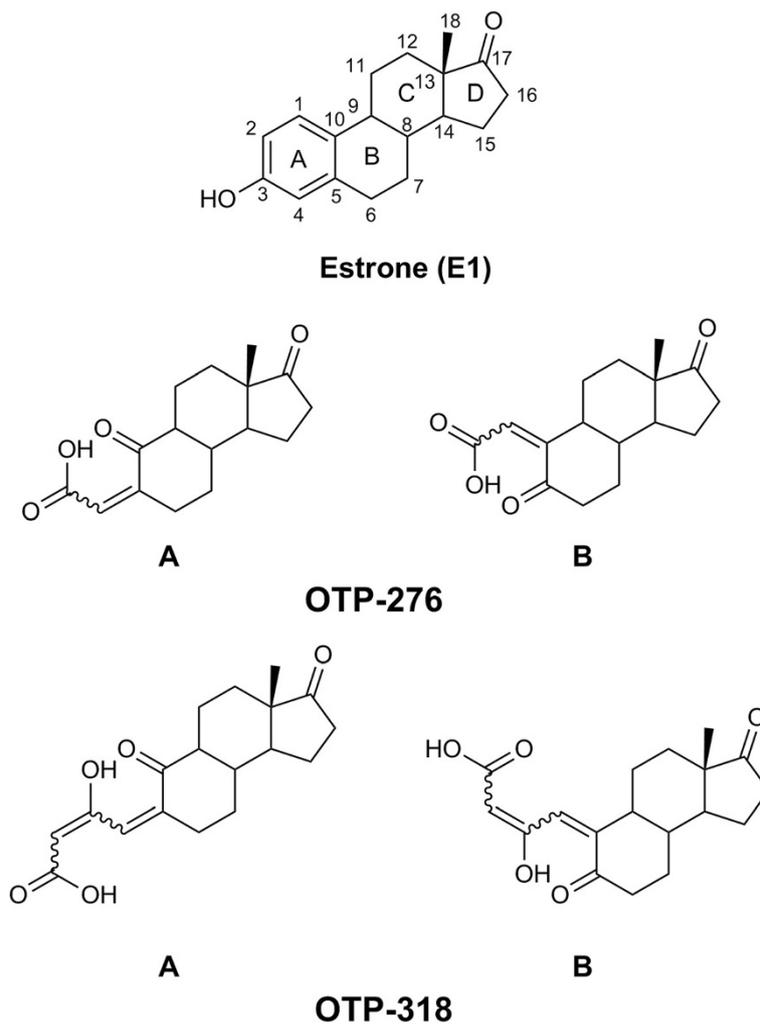


Figure 1 Molecular structure of estrone (E1) showing the standard C atom numbering and two possible structures for each of the identified OTPs. OTP-276B was not observed, while OTP-318B was 10 times less abundant than OTP-318A. Wavy bonds indicate that the exact stereochemistry is unknown.

partially successful (only one compound was found, with a precursor ion at $m/z = 275$ which will be discussed later). For this reason, a control compare trend experiment, followed by differential analysis with the Sieve software was performed. Sieve found 593 unique “frames” in the samples. In this software, a frame represents the coordinates of a peak in a three-dimensional space that allows the identification of substances in the HRMS acquisition files. The coordinates of these frames are: retention time, m/z value and intensity. However, not all of these frames represent an OTP of E1. Many frames correspond to background signals and laboratory contaminants that are detected by the mass spectrometer. In order to eliminate the frames that are not relevant to the identification of OTPs of E1, the frames table filter function available in Sieve was used. This feature allows to set several rules that the frames must follow in order to qualify as a potential OTP. The rules are based on parameters such as minimum or maximum m/z value, retention time (t_R) range, average frame intensity, normalized average frame intensity to control (samples E1:O₃ 1:8), etc. We applied a set of three rules to identify the frames most probably corresponding to OTPs of E1: i) The normalized average intensity of a frame in the samples E1:O₃ 1:0 must be < 0.05 ; ii) Normalized average intensity of a frame in samples E1:O₃ 1:1 and 1:5 > 0.05 ; and iii) The average intensity of the frame in the samples E1:O₃ 1:8 must be > 5000 . The first rule ensured that frames present in the sample E1:O₃ 1:0, which was not ozonated, had very low intensity compared to the intensity in the sample E1:O₃ 1:8, which had the largest dose of O₃. Normally a frame corresponding to an OTP should not be observed in the sample E1:O₃ 1:0 but since some low carryover was observed between samples, residual ultra trace amounts were often detected in all the samples. High carryover could be problematic as it can mask significant differences between the samples. Therefore it was kept to a minimum by applying syringe washes and using blanks between samples. The second rule guaranteed that the filtered frames are not associated to random signal spikes since they have to be present in both samples E1:O₃ 1:0 and 1:5. Therefore their normalized average intensity has to be different than zero. The normalized value of 0.05 was found to be optimal to eliminate most of frames containing peaks with unacceptable shape. The last rule ensured that only frames of relevant intensity are kept for further analysis since frames an average intensity of < 5000 were not considered as main OTPs. This rule also helped selecting frames with intensity sufficiently high to allow MSⁿ experiments. The filtered frame results are summarized in Table 1. It can be noted that only 16 out of the 593 initial frames were conserved and interestingly only two (frame #9 and #78) were among the 100 most intense frames. Visual examination of the reconstructed ion chromatograms showed that only these

two frames (#9 and #78, eluting at $t_R = 3.51$ and 3.32 min, respectively) had acceptable peak shape. The others had peaks with retention times that were not consistent across all the samples or had low signal-to-noise ratios. Looking at the retention times of the frames after the filter was applied, it can be seen that many of the frames identified by the differential analysis software as OTPs eluted by the end of the chromatographic run ($t_R > 7.0$ min). Inspection of these frames showed that they consisted mostly of intense noise with signal spikes and is unlikely that they were related to E1 (it is not clear, however, why these frames were not eliminated by the filters). Lower retention times than the parent compound are consistent with OTPs which by the addition of O and loss of C atoms tend to be more hydrophilic than the parent compound (t_R of E1 = 4.87 min) and thus less retained in reversed-phase columns. Therefore only frames #9 and #78 were kept for further investigation.

The elemental compositions of the OTPs present in frame #9 and #78, presented in Table 2, were determined by QualBrowser according to the parameters mentioned in the *Identification of the main ozonation transformation products of estrone* section. Since these compounds are a result of the oxidation of E1, which has an elemental composition of C₁₈H₂₂O₂, the number of elements and their constraints used for generating formulas with the Elemental Composition function in QualBrowser significantly reduced the number of possible matches. Given that after ozonolysis the number of C atoms in the OTPs is not expected to increase, the maximum number of C atoms was set to 18 (minimum was always set to 0). As for O, a value higher than 10 is not expected given that the number of reaction sites is limited (phenol function in ring A and ketone at C-17, see Figure 1). Finally for H, a maximum value of 40 was set as ring cleavage caused by O₃ could lead to the formation of new functional groups containing H such as -COOH, -CHO and -OH [34,35]. Also in order to reduce the number of possible matches, the nitrogen rule and a mass tolerance of 10 mmu were employed. The software returned only one match for the composition of the OTP in frame #9, C₁₆H₁₉O₄ (C₁₆H₂₀O₄ for the neutral molecule) with a $\Delta m_{mu} = 0.418$. This composition implies that the reaction of E1 with O₃ caused the loss of 2 C atoms and 2 H atoms as well as the addition of 2 O atoms. As for the OTP in frame #78, two elemental compositions were suggested. The most likely composition was C₁₈H₂₁O₅ (addition of 3 O atoms with respect to E1) due to its RDBE value of 8.5 (same as that of E1). The other proposed composition, C₁₁H₂₅O₁₀ (RDBE = -0.5), was a fully saturated compound that is a very unlikely possibility given that the pseudo-molecular ion of E1 has a RDBE = 8.5 (four rings plus four π bonds, the 0.5 value is due to the loss of a H atom in the [M-H]⁻ ion).

Table 1 Characteristics of the frames retained for further analysis by applying the filter option

Frame number ^a	<i>m/z</i>	Retention time (min)	Normalized average intensities ^b			Average intensity in sample 1:8
			Sample 1:0	Sample 1:1	Sample 1:5	
9	275.12930	3.51	0.009	0.062	0.614	22664
78	317.14008	3.32	0.022	0.075	0.914	5323
200	212.09456	7.38	0.050	0.486	0.863	8552
225	224.06932	7.49	0.026	0.442	0.739	6080
241	202.07396	7.33	0.049	0.483	0.926	11688
267	224.09468	7.44	0.036	0.456	0.916	7829
282	228.08983	7.40	0.039	0.297	0.694	6061
294	226.07419	7.46	0.000	0.456	0.943	6511
333	226.08574	7.40	0.000	0.417	0.652	6489
344	229.08455	7.41	0.049	0.334	0.540	5285
357	212.05852	7.46	0.041	0.466	0.784	5656
358	201.07838	7.45	0.041	0.477	0.869	7736
375	215.06937	7.46	0.044	0.451	0.566	5559
418	250.14478	4.29	0.038	0.071	0.239	9056
419	250.14525	3.70	0.027	0.086	0.303	8340
574	250.14519	2.99	0.038	0.051	0.240	8590

^a Frame number is assigned sequentially by the software according to absolute intensity in all samples. Therefore frame #1 represents the most intense frame found across all samples. ^b Average intensity of each frame was normalized to its average intensity in the samples to the control (sample 1:8).

The results showed that the differential analysis method significantly reduces the time of data analysis and interpretation. Background noise, sample contaminants and signal spikes were thus mostly eliminated from the files by using a control and series of samples with decreasing concentration of O₃ as well as a set of rules to filter the results. Therefore instead of performing manual analysis (elemental composition, acceptable peak shape) of 593 potential OTPs, this method reduced the data set to 16 compounds. While this method still suffers from an abundance of false positive hits (only 12.5% of the potential OTPs were good candidates) and may also ignore minor OTPs, it proved to be a rapid and simple way to identify the two main OTPs of E1 formed during the ozonation conditions described in the *Ozonation of estrone samples* section.

Structural elucidation of OTPs

Preconcentration of ozonated samples was necessary in order to gain more information about the molecular structure of the identified OTPs by performing MSⁿ experiments. If concentrations of OTPs would have been sufficiently high in the ozonated samples, data-dependent experiments could have been performed to obtain additional structural information at the identification stage. However, in this study a SPE step was necessary in order to obtain signal intensities high enough for reliable MS²ⁿ experiments. Injection of the SPE extract of the E1 ozonated solutions showed intense chromatographic peaks eluting at

*t*_R = 3.52 min and 3.40 min with mass spectra showing base peaks at *m/z* 275.1 and *m/z* 317.1, respectively. These results confirmed that the OTP present in frame #9 (called OTP-276 from now on) and frame #78 (called OTP-318) were successfully preconcentrated by SPE. Interestingly, two other peaks were observed with *t*_R=3.64 and *t*_R=3.73 min at *m/z* 317.14 ± 0.01, which may imply that isomers of the compound were also formed (Additional file 1: Figure S1). Direct infusion of these extracts into the mass spectrometer allowed performing multi-stage tandem mass spectrometry (MSⁿ) experiments with these ions. It is worth mentioning that the structural elucidation stage of the present method can be further simplified by using software capable of predicting collision-induced product ions [36]. This approach was unfortunately not possible here since the software available to us was not able to predict product ions of even-electron negative ions.

Table 2 Elemental composition of the two potential OBPs identified by the differential analysis software compared to that of the parent compound E1

Frame number	<i>m/z</i>	Elemental composition	RDBE	Δ mmu
(E1)	269.15537	C ₁₈ H ₂₁ O ₂	8.5	0.667
9	275.12930	C ₁₆ H ₁₉ O ₄	7.5	0.418
78	317.14008	C ₁₈ H ₂₁ O ₅	8.5	0.633
		C ₁₁ H ₂₅ O ₁₀	-0.5	-5.240

RDBE: Ring and double bond equivalents. Δmmu: absolute mass accuracy in millimass units.

Structural elucidation of OTP-276 by HRMSⁿ experiments

Ozone is a very selective oxidant towards electron-rich groups [37] and the E1 molecule has only two of such groups, the aromatic ring (ring A) and the ketone function at C-17 (Figure 1). Of these two, the most likely site of reaction is the aromatic ring since it has higher electron density, especially in the *ortho* and *para* positions to the electron-donating hydroxyl group at C-3 [35]. As determined in the previous section, the elemental composition of OTP-276 is C₁₆H₁₉O₄, which indicates the loss of 2 C atoms and 2 H atoms as well as the addition of 2 O atoms with respect to E1. Fragmentation of the pseudo-molecular ion of OTP-276 (*m/z* 275) by collision-induced dissociation (CID) with 30 normalized collision energy (NCE) in the linear ion trap portion of the mass spectrometer yielded only one main first generation (MS²) product ion, *m/z* 231.13989 with the elemental composition C₁₅H₁₉O₂ ($\Delta m_{mu}=0.837$). The loss of 44 u with respect to the precursor ion indicates the loss of CO₂ by heterolytic cleavage. Decarboxylation is a characteristic fragmentation pathway for even-electron negative ions of carboxylic acids [32,38]. Therefore a carboxylic acid functional group must be present in the structure of OTP-276. This functional group could have been formed after ring cleavage and loss of two aromatic C atoms of E1 to form a ketone group on C-10 and a carboxylic acid on C-3 (OTP-276A, in Figure 1) after ozonolysis. This structure was previously suggested by de Oliveira Pereira, et al. [14]. It was explained by a two-step O₃ attack, the first forming a molozonide intermediate yielding an aldehyde and a carboxylic acid on C-2 and C-3 respectively, and the second forming a new molozonide intermediate that decomposes to form OTP-276 [14]. When performing a full scan (MS¹) experiment on the SPE extract of ozonated E1-d₄, the formation of an isomer of this structure having ketone group on C-5 and the carboxylic group on C-2 (OTP-276B, Figure 1) with a formula C₁₆H₁₇D₂O₄ was not observed. The ion at *m/z* 278.14758 (C₁₆H₁₆D₃O₄, $\Delta m_{mu}=0.964$) was observed instead; indicating that one of the aromatic D atoms was still present in the molecular structure. This experiment thus reinforces the validity of the proposed structure for (OTP-276A, Figure 1).

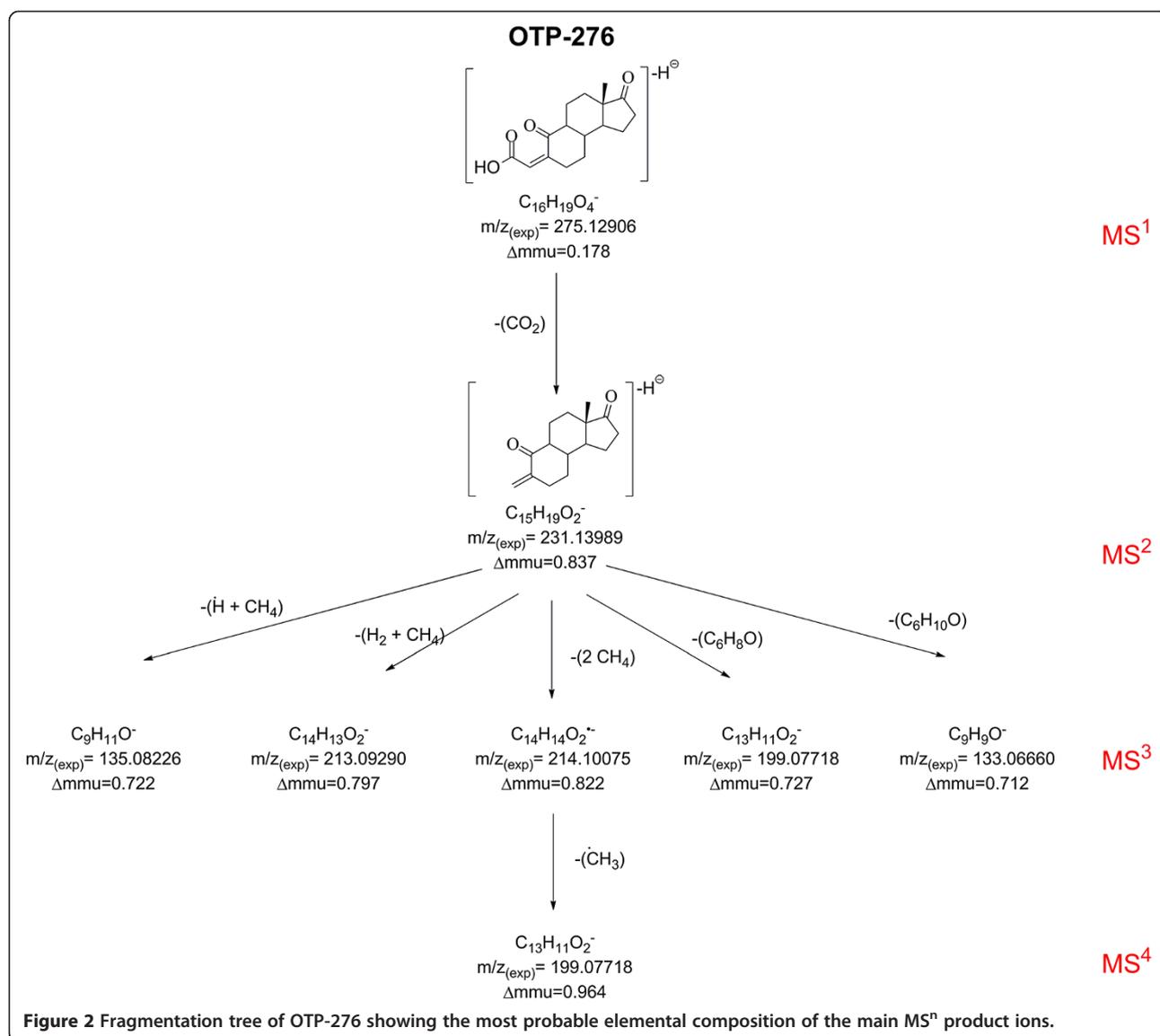
Following these results, MSⁿ (*n*=3-4) experiments were performed with the precursor ion at *m/z* 275 in order to further confirm the proposed molecular structure of OTP-276. The fragmentation tree of *m/z* 275 showing the formulas of the observed MSⁿ product ions can be seen in Figure 2. The presence of the second generation (MS³) product ion at *m/z* 214 is particularly puzzling since it does not follow the nitrogen rule. Therefore this ion must be odd-electron, meaning that *m/z* 214 violates the even-electron rule which states that even-electron ions do not lose a radical to form an odd-electron ion [39]. However, such deviations of the even-electron rule have been

previously reported after CID of even-electron ions [38,40]. The loss of 17 u was thus interpreted as the loss of H⁺ + CH₄. The minor MS³ product ions observed in the full scan CID spectrum of *m/z* 231 at *m/z* 213.09290 (C₁₄H₁₃O₂, $\Delta m_{mu}=0.797$), *m/z* 199.07718 (C₁₃H₁₁O₂, $\Delta m_{mu}=0.727$), *m/z* 135.0826 (C₉H₁₁O[•], $\Delta m_{mu}=0.722$) and *m/z* 133.06660 (C₉H₉O[•], $\Delta m_{mu}=0.712$) indicate neutral losses of CH₆, C₂H₈, C₆H₈O and C₆H₁₀O, respectively. Since the loss H₂ and CH₄ is a common pattern of fragmentation for negative even-electron ions [32], the neutral loss of CH₆ was interpreted as the combined loss H₂ + CH₄, and that of C₂H₈ as 2CH₄. Losses of CH₄ can be explained by the dissociation in stepwise elimination mechanisms [32] of the methyl group from C-13 (the C atom numbering of E1 is preserved to facilitate the identification of the fragmentation sites) and the methylene group from C-5. However, the precise mechanism that could yield H₂ and CH₄ losses from *m/z* 231 remains unclear. Additionally, the formation of ions at *m/z* 135 and *m/z* 133 is also difficult to elucidate, however they are likely the consequence of stepwise elimination reactions and internal nucleophilic displacements [32]. The elemental compositions of the second generation product ions formed during the MS³ experiments were confirmed using the E1-d₄ ozonated SPE extract (Additional file 1: Figure S2). The results of these experiments showed that a D atom was lost during the fragmentation of *m/z* 234.15759 (C₁₅H₁₆D₃O₂, $\Delta m_{mu}=-0.293$) to form *m/z* 201.08897 (C₁₃H₉D₂O₂, $\Delta m_{mu}=-0.246$). This D atom was most likely located in the methylene group on C-5. The third generation (MS⁴) product ion scan experiment (*m/z* 275 → *m/z* 231 → *m/z* 214 → *m/z* 50–300) with NCE=50 resulted in the presence of only one ion at *m/z* 199.07631. This third generation product ion was likely formed by dissociation of [•]CH₃.

In summary, MSⁿ experiments data showed that only the MS² product ion at *m/z* 231 clearly demonstrates the presence of a carboxylic acid group in the OTP-276 structure. The ion at *m/z* 199 observed in the MS³ and third generation (MS⁴) product ion scans could have suggested the presence of a double bond between C-4 and C-5 in OTP-276, however the cleavage of this bond to yield the CH₄ or ([•]CH₃) neutral losses observed was not easily explained by known mechanisms. The rest of the neutral losses, albeit characteristic of negative even-electron ions, did not suggest the presence of other functional groups on the molecule. Product ions generated in the MS³ to MS⁴ experiments pointed towards a partial preservation of the ring structure of E1 and the observed losses (H₂, CH₄) were thus justified by the formation of resonance-stabilized ring structures.

Structural elucidation of OTP-318 by HRMSⁿ experiments

Fragmentation of the precursor ion at *m/z* 317 resulted in the formation of a MS² product ion at *m/z* 273.14960.



The neutral loss (44 u) corresponds to CO_2 , which as in the case of OTP-276, indicates the presence of a carboxylic acid group in the molecular structure of OTP-318. Initially it was thought that the structure of OTP-318 was 1,2,4-trihydroxyestrone (hydroxylation at C-1, C-2 and C-4 of E1). This structure was rejected once it was observed that a CO_2 loss was difficult to originate from such a compound. The carboxylic group in OTP-318 was most likely formed after the cleavage of the aromatic ring of E1 by O_3 . This was confirmed by the full scan spectrum of the ozonated extract of E1- d_4 (Additional file 1: Figure S3), which yielded an abundant ion at m/z 321.16456 ($C_{16}H_{17}D_4O_5^-$, $\Delta mmu = 0.006$). Since the deuterated OTP conserved all its D atoms after ozonolysis, the O_3 attack must have occurred at C-1 and C-10 rather than at C-4 and C-5, which would result in the loss of the D atom at C-4 (OTP-318B, Figure 1). Interestingly

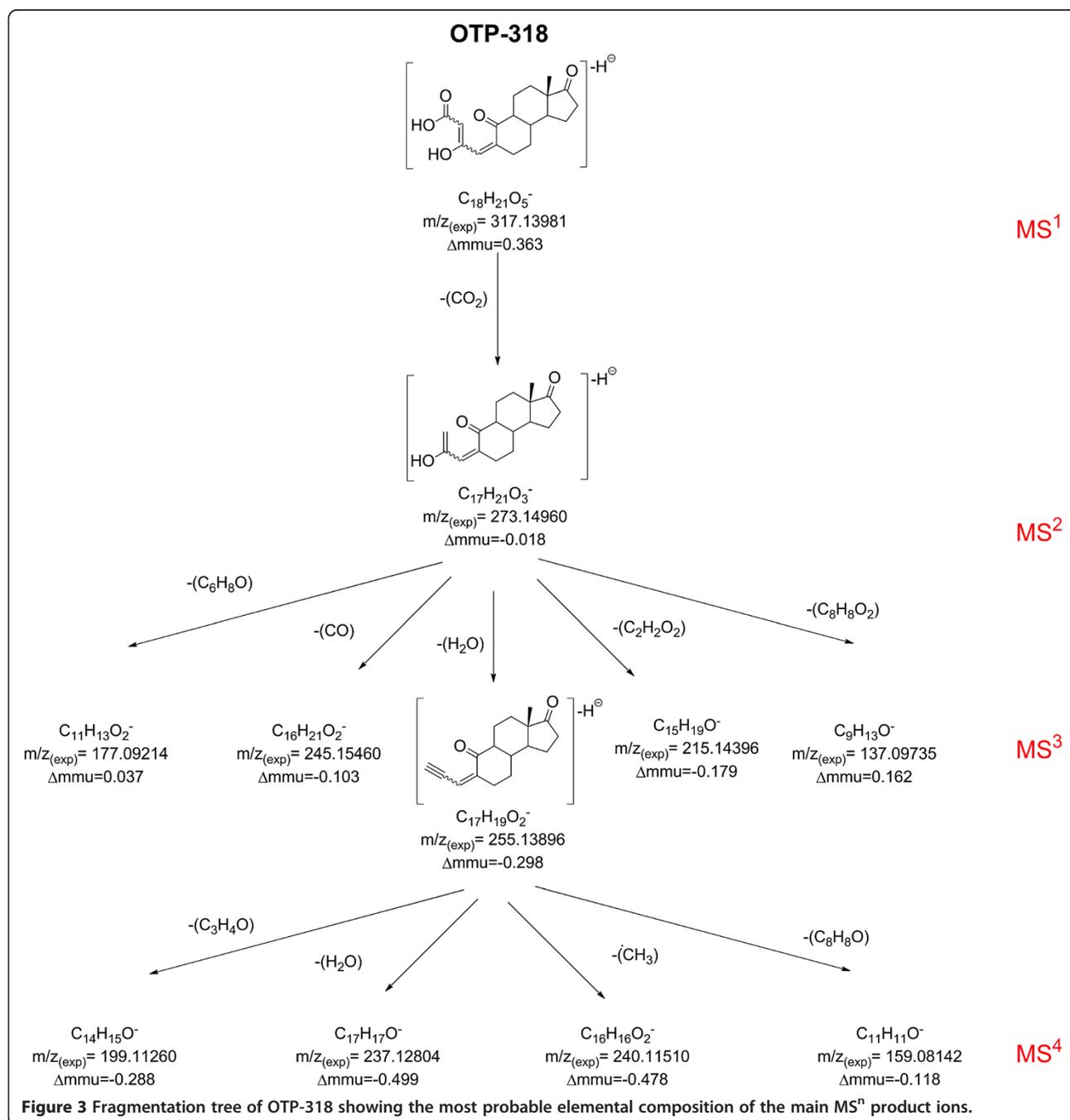
the formation of an OTP-318 isomer resulting from the O_3 attack at C-4 and C-5 ($C_{18}H_{18}D_3O_5^-$, $\Delta mmu = -0.217$) was also observed in the mass spectrum at m/z 320.15806, but its signal was less intense, about 10 times less abundant than m/z 321.

MS³ and MS⁴ experiments were performed on the most abundant ions in order to gain more information about the structure of OTP-318. The MS³ product ion scan (m/z 317 → m/z 273 ↗ m/z 75–350) showed the presence of five peaks with abundance > 10%. These were: m/z 255.13896 ($C_{17}H_{19}O_2^-$, $\Delta mmu = -0.298$); m/z 245.15460 ($C_{16}H_{21}O_2^-$, $\Delta mmu = -0.103$), m/z 215.14396 ($C_{15}H_{19}O^-$, $\Delta mmu = -0.179$), m/z 177.09214 ($C_{11}H_{13}O^-$, $\Delta mmu = 0.037$) and m/z 137.09735 ($C_9H_{13}O^-$, $\Delta mmu = 0.162$). The loss of H_2O (18 u) that yields m/z 255 is rather confusing since this type of neutral loss is not very frequent in negative even-electron ions [38]. However, loss of H_2O was

interpreted as the result of the formation of an alkynyl group after rupture of the C-OH bond at C-3 in OTP-318. A similar type of mechanism has been proposed during the fragmentation of ethers [41]. Observation of MS³ product ion scan spectrum of the E1-d₄ ozonated SPE extract showed the presence of two ions which could be explained by this type of loss: *m/z* 259.16380 (C₁₇H₁₅D₄O₂⁻, Δ_{mmu}=0.737) and *m/z* 258.15754 (C₁₇H₁₆D₃O₂⁻, Δ_{mmu}=-0.343). The elemental composition of these two MS³ products ions thus seems

to indicate that the loss of H₂O or HDO is the result of the formation of a hydroxide anion complex.

The rest of the product ions do not indicate clearly the presence of other functional groups in the OTP-318 structure. The MS⁴ product ion scan (*m/z* 317 → *m/z* 273 → *m/z* 255 ↗ *m/z* 75–350) yielded four main ions: *m/z* 240.11510 (C₁₆H₁₆O₂⁻, Δ_{mmu}=-0.478), *m/z* 237.12804 (C₁₇H₁₇O⁻, Δ_{mmu}=-0.499), *m/z* 199.11260 (C₁₄H₁₅O⁻, Δ_{mmu}=-0.288) and *m/z* 159.08142 (C₁₁H₁₁O⁻, Δ_{mmu}=-0.118). From these the ion at *m/z* 237 suggested a second



loss of H₂O but it is not clear from which O atom (ketone group at C-10 or C-17) this loss was originated. As it was the case with OTP-276, the presence of *m/z* 240 in the MS⁴ product ion scan points towards the formation of an odd-electron ion, most likely from the loss of •CH₃ from C-13.

In summary, MSⁿ experiments with the ozonated extracts of E1 and E1-d₄ showed that a carboxylic acid and a hydroxyl group are present in the OTP-318 molecule. The other fragments showed in Figure 3 did not indicate unambiguously the presence of other functional groups in the molecular structure of OTP-318. Similarly to the fragments observed during the MSⁿ experiments performed with OTP-276, the elemental composition of the MS³ and MS⁴ product ions of OTP-318 pointed towards a partial preservation of the ring structure of E1. However, typical negative even-electron losses such as H₂ and CH₄ were observed less frequently than in OTP-276, probably due to the presence of 2 more O atoms in the structure OTP-318 parent ion which could have an impact on the possible fragmentation mechanisms. The proposed structures in Figure 1 (OTP-318A and OTP-318B) also hint that at least 4 configuration isomers are probable for this molecule given the presence of two carbon double bonds in its structure. This could explain the observation of at least three peaks at *m/z* 317.14 ± 0.01 in the chromatogram of the SPE extracts of the ozonated E1 solutions (Additional file 1: Figure S1).

Conclusions

The present work showed that the identification, structural elucidation and confirmation of OTPs of contaminants of emerging concern can be significantly simplified using a control compare trend experiment followed by differential analysis and MSⁿ experiments. Application of this method to the study of the OTPs of E1 allowed the identification of two compounds of mass 276 u (OTP-276) and 318 u (OTP-318). Structural elucidation of these compounds was performed by MSⁿ (*n*=2-4) experiments via direct infusion into the mass spectrometer of SPE extracts of ozonated solutions of E1 and E1-d₄. Observation of a neutral loss corresponding to CO₂ in the MS² product ion scan indicated the presence of a carboxylic acid group in the molecular structure of both OTPs. Also a loss of H₂O observed in the MS³ product ion scan of OTP-318 suggested the presence of a hydroxyl group. The presence of these functional groups is consistent with known reaction mechanisms of O₃ [14,35] and previously suggested OTPs of hormones [11,12,15,27]. A study by de Oliveira Pereira, et al. [14] also identified OTP-276 as a main by-product of the ozonation of E1. However de Oliveira Pereira, et al. [14] and Huber, et al. [11] also found other OTPs that were not identified in the samples using the method

presented in this paper. This might be a consequence of the different ozonation setups and experimental conditions used during the ozonation process.

While the method proposed here might oversight minor OTPs, it proved to be successful to eliminate most of the background noise, sample contaminants and signal spikes that are present in the acquisition files and to streamline the identification of the main OTPs. Removal of all the irrelevant data reduced to a great extent the amount of information that had to be processed manually (from 593 to 16 frames). Future work will focus on the optimization of software parameters to limit the number of candidate frames and reduce false positives. This method can now be more widely applied to the identification and elucidation of OTPs of other contaminants of emerging concern such as illicit drugs and antibiotics.

Additional file

Additional file 1: Figure S1 . LC-HRMS chromatogram of the ozonated E1 pre-concentrated sample. **Figure S2**. Fragmentation tree of OTP-279 (deuterated analogue of OTP-276). **Figure S3**. Fragmentation tree of OTP-322 (deuterated analogue of OTP-318).

Abbreviations

CECs: Contaminants of emerging concern; CID: Collision-induced dissociation; E1: Estrone; E1-d₄: Deuterium-labeled estrone; GC-MS: Gas chromatography–mass spectrometry; HRMS: High-resolution mass spectrometry; LC-MS/MS: Liquid chromatography–tandem mass spectrometry; MSⁿ: Multi-stage tandem mass spectrometry; NCE: Normalized collision energy; OTPs: Ozonation transformation products; RDBE: Ring and double bonds equivalents; TPs: Transformation products; t_R: Retention time.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

PAS participated in the conception and design of the experiments, in the acquisition, analysis and interpretation of data and in the drafting and revising of the manuscript. PK collaborated to the sample preparation and data analysis and also revised the manuscript. VY contributed to the design of the experiments, to the interpretation of the data and corrected the manuscript. All authors read and approved the final manuscript.

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References

1. Celiz MD, Tso J, Aga DS: **Pharmaceutical metabolites in the environment: analytical challenges and ecological risks.** *Environ Toxicol Chem* 2009, **28**:2473–2484.
2. Andreozzi R, Caprio V, Ciniglia C, De Champdore M, Lo Giudice R, Marotta R, Zuccato E: **Antibiotics in the environment: occurrence in Italian STPs, fate, and preliminary assessment on algal toxicity of amoxicillin.** *Environ Sci Technol* 2004, **38**:6832–6838.
3. Boreen AL, Arbold WA, McNeill K: **Photodegradation of pharmaceuticals in the aquatic environment: a review.** *Aquat Sci* 2003, **65**:320–341.

4. Yargeau V, Leclair C: **Impact of operating conditions on decomposition of antibiotics during ozonation: a review.** *Ozone Sci Eng* 2008, **30**:175–188.
5. Quintana JB, Weiss S, Reemtsma T: **Pathways and metabolites of microbial degradation of selected acidic pharmaceutical and their occurrence in municipal wastewater treated by a membrane bioreactor.** *Water Res* 2005, **39**:2654–2664.
6. Boxall AB, Sinclair CJ, Kolpin D, Maund SJ: **When synthetic chemicals degrade in the environment.** *Environ Sci Technol* 2004, **38**:368A–375A.
7. Horn O, Nalli S, Cooper D, Nicell J: **Plasticizer metabolites in the environment.** *Water Res* 2004, **38**:3693–3698.
8. Boxall A, Rudd M, Brooks B, Caldwell D, Choi K, Hickmann S, Innes E, Ostapyk K, Staveley J, Verslycke T, Ankley G, Beazley K, Belanger S, Berninger J, Carriquiriborde P, Coors A, Deleo P, Dyer S, Ericson J, Gagné F, Giesy J, Gouin T, Hallstrom L, Larsson M, Larsson D, Lazorchak J, Mastrocco F, McLaughlin A, McMaster M, Meyerhoff R, Moore R, Parrott J, Snape J, Murray-Smith R, Servos M, Sibley P, Straub J, Szabo N, Topp E, Tetreault G, Trudeau V, Van Der Kraak G: **Pharmaceuticals and personal care products in the environment: what are the Big questions?** *Environ Health Perspect* 2012, **120**:1221–1229.
9. Kosjek T, Heath E: **Applications of mass spectrometry to identifying pharmaceutical transformation products in water treatment.** *TRAC Analyt Chem* 2008, **27**:807–820.
10. Ikehata K, Naghashkar NJ, El-Din MG: **Degradation of aqueous pharmaceuticals by ozonation and advanced oxidation processes: A review.** *Ozone Sci Eng* 2006, **28**:353–414.
11. Huber MM, Ternes TA, Von Gunten U: **Removal of estrogenic activity and formation of oxidation products during ozonation of 17 α -ethynylestradiol.** *Environ Sci Technol* 2004, **38**:5177–5186.
12. Bila D, Montalvão AF, Azevedo DA, Dezotti M: **Estrogenic activity removal of 17 β -estradiol by ozonation and identification of by-products.** *Chemosphere* 2007, **69**:736–746.
13. Radjenović J, Godehardt M, Hein A, Farré M, Jekel M, Barceló D: **Evidencing generation of persistent ozonation products of antibiotics roxithromycin and trimethoprim.** *Environ Sci Technol* 2009, **43**:6808–6815.
14. de Oliveira PR, Lopez de Alda M, Joglar J, Daniel LA, Barceló D: **Identification of new ozonation disinfection byproducts of 17 [beta]-estradiol and estrone in water.** *Chemosphere* 2011, **84**:1535–1541.
15. Larcher S, Delbès G, Robaire B, Yargeau V: **Degradation of 17 α -ethynylestradiol by ozonation—Identification of the by-products and assessment of their estrogenicity and toxicity.** *Environ Int* 2012, **39**:66–72.
16. Benner J, Ternes T: **Ozonation of propranolol: formation of oxidation products.** *Environ Sci Technol* 2009, **43**:5086–5093.
17. Kern S, Fenner K, Singer HP, Schwarzenbach RP, Hollender J: **Identification of transformation products of organic contaminants in natural waters by computer-aided prediction and high-resolution mass spectrometry.** *Environ Sci Technol* 2009, **43**:7039–7046.
18. Helbling DE, Hollender J, Kohler H-PE, Singer H, Fenner K: **High-throughput identification of microbial transformation products of organic micropollutants.** *Environ Sci Technol* 2010, **44**:6621–6627.
19. Gómez-Ramos MM, Pérez-Parada A, García-Reyes JF, Fernández-Alba AR, Agüera A: **Use of an accurate-mass database for the systematic identification of transformation products of organic contaminants in wastewater effluents.** *J Chromatogr A* 2011, **1218**:8002–8012.
20. Gervais G, Bichon E, Antignac J-P, Monteau F, Leroy G, Barriatua L, Chachignon M, Ingrand V, Roche P, Bizac BL: **Differential global profiling as a new analytical strategy for revealing micropollutant treatment by-products: Application to ethynylestradiol and chlorination water treatment.** *Chemosphere* 2011, **83**:1553–1559.
21. Servos M, Bennie D, Burnison B, Jurkovic A, McInnis R, Neheli T, Schnell A, Seto P, Smyth S, Ternes T: **Distribution of estrogens, 17 β -estradiol and estrone, in Canadian municipal wastewater treatment plants.** *Sci Total Environ* 2005, **336**:155–170.
22. Kolpin DW, Furlong ET, Meyer MT, Thurman EM, Zaugg SD, Barber LB, Buxton HT: **Pharmaceuticals, hormones, and other organic wastewater contaminants in U.S. streams, 1999–2000: a national reconnaissance.** *Environ Sci Technol* 2002, **36**:1202–1211.
23. Benotti M, Trenholm R, Vanderford B, Holady J, Stanford B, Snyder S: **Pharmaceuticals and endocrine disrupting compounds in US drinking water.** *Environ Sci Technol* 2009, **43**:597–603.
24. Snyder SA, Wert EC, Rexing DJ, Zegers RE, Drury DD: **Ozone oxidation of endocrine disruptors and pharmaceuticals in surface water and wastewater.** *Ozone Sci Eng* 2006, **28**:445–460.
25. Hart EJ, Sehested K, Holoman J: **Molar absorptivities of ultraviolet and visible bands of ozone in aqueous solutions.** *Anal Chem* 1983, **55**:46–49.
26. Paraskeva P, Lambert S, Graham N: **Influence of ozonation conditions on the treatability of secondary effluents.** *Ozone Sci Eng* 1998, **20**:133–150.
27. Vieira K, Nascentes C, Augusti R: **Ozonation of ethynylestradiol in aqueous-methanolic solution: direct monitoring by electrospray ionization mass spectrometry.** *J Braz Chem Soc* 2010, **21**:787–794.
28. Keller BO, Sui J, Young AB, Whittall RM: **Interferences and contaminants encountered in modern mass spectrometry.** *Anal Chim Acta* 2008, **627**:71–81.
29. Rodayan A, Roy R, Yargeau V: **Oxidation products of sulfamethoxazole in ozonated secondary effluent.** *J Hazard Mater* 2010, **177**:237–243.
30. Shareef A, Angove MJ, Wells JD, Johnson BB: **Aqueous solubilities of estrone, 17 β -estradiol, 17 α -ethynylestradiol, and bisphenol A.** *J Chem Eng Data* 2006, **51**:879–881.
31. Segura PA, MacLeod SL, Lemoine P, Sauvé S, Gagnon C: **Quantification of carbamazepine and atrazine and screening of suspect organic contaminants in surface and drinking waters.** *Chemosphere* 2011, **84**:1085–1094.
32. Bowie JH: **The fragmentations of even-electron organic negative ions.** *Mass Spectrom Rev* 1990, **9**:349–379.
33. McLafferty FW, Tureček F: **Basic Mechanisms of Ion Fragmentation.** In *Interpretation of Mass Spectra*. Sausalito, CA: University Science Books; 1993:51–84.
34. Yamamoto Y, Niki E, Shiokawa H, Kamiya Y: **Ozonation of organic compounds. 2. Ozonation of phenol in water.** *J Org Chem* 1979, **44**:2137–2142.
35. Decoret C, Royer J, Legube B, Dore M: **Experimental and theoretical studies of the mechanism of the initial attack of ozone on some aromatics in aqueous medium.** *Environ Technol Lett* 1984, **5**:207–218.
36. Segura PA, Gagnon C, Yargeau V: **On the application of high-resolution mass spectrometry to environmental analysis.** *Int J Env Poll Remed* 2012, **1**:85–92.
37. Oyama ST: **Chemical and catalytic properties of ozone.** *Catal Rev* 2000, **42**:279–322.
38. Levsen K, Schiebel HM, Terlouw JK, Jobst KJ, Elend M, Preiß A, Thiele H, Ingendoh A: **Even-electron ions: a systematic study of the neutral species lost in the dissociation of quasi-molecular ions.** *J Mass Spectrom* 2007, **42**:1024–1044.
39. Gross JH: **Fragmentation of Organic Ions and Interpretation of EI Mass Spectra.** In *Mass Spectrometry: A Textbook*. Berlin, Germany: Springer; 2004:223–330.
40. Thurman EM, Ferrer I, Pozo OJ, Sancho JV, Hernandez F: **The even-electron rule in electrospray mass spectra of pesticides.** *Rapid Commun Mass Spectrom* 2007, **21**:3855–3868.
41. Eichinger PCH, Bowie JH: **Gas-phase carbanion rearrangements. Deprotonated benzyl and allyl ethers.** *J Chem Soc, Perkin Trans II* 1988, **4**:497–506.

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