

Poster presentation

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Selectivity of new caspase 3 and 8 tetrapeptide substrates can be explained by automated docking analysis

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from 4th German Conference on Chemoinformatics
Goslar, Germany. 9–11 November 2008

Published: 5 June 2009

Chemistry Central Journal 2009, 3(Suppl 1):P76 doi:10.1186/1752-153X-3-S1-P76

This abstract is available from: <http://www.journal.chemistrycentral.com/content/3/S1/P76>

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Caspases are cysteine proteases and are considered key-mediators in apoptotic cell death. Selective quantification of the various caspase activities in cancer cells is important for detecting cell death caused by cancer therapy. We have designed and synthesized a series of novel fluorogenic tetrapeptide substrates for caspase 8 and investigated the substrates for selective cleavage by either caspases 3 or 8 in enzyme assays. At the same time we have used the automated docking program AutoDock (ver 3, [1,2]) to dock the new substrates into the active sites of X-ray crystal structures of human caspases 3 and 8, respectively. AutoDock was confirmed to be an appropriate tool for substrate binding prediction because substrate docking results are comparable with documented X-ray crystal structure of caspase 3 and 8 bounded with analogous tetrapeptide inhibitors [3,4]. Enzyme-substrate conformations with changes in free energy of binding (ΔG) were calculated with AutoDock and compared to the experimental determined Michaelis-Menten constant K_m . A significant correlation between the experimental K_m and theoretical ΔG was found. Enzyme kinetics showed the substrates to have 100-fold lower K_m -values for caspase 8 compared to caspase 3. This selectivity was reflected in the significantly larger negative ΔG -values between the substrates docked to caspase 8 as opposed to caspase 3. These results will help in the design of even more selective caspase substrates.

References

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